LXR deficiency and cholesterol feeding affect the expression and phenobarbital-mediated induction of cytochromes P450 in mouse liver

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Abstract Metabolic transformation by the superfamily of cytochromes P450 (CYPs) plays an important role in the detoxification of xenobiotics such as drugs, environmental pollutants, and food additives. Endogenous substrates of CYPs include fatty acids, sterols, steroids, and bile acids. Induction of CYPs via transcriptional activation by substrates and other xenobiotics is an important adaptive mechanism that increases the organism's defense capability against toxicity. Numerous in vivo and in vitro data have highlighted the concept that the molecular mechanism of hepatic drug induction is linked to endogenous regulatory pathways. In particular, in vitro data suggest that oxysterols via the liver X receptor (LXR) inhibit phenobarbital (PB)-mediated induction of CYPs. To study the link between LXR, cholesterol homeostasis, and drug induction in vivo, we designed experiments in wild-type, LXR−/−, LXRα−/−, and LXRβ−/−/β-deficient mice. Our data expose differential regulatory patterns for Cyp2b10 and Cyp3a11 dependent on the expression of LXR isoforms and on challenge of cholesterol homeostasis by excess dietary cholesterol. Our results suggest that, in the mouse, liver cholesterol status significantly alters the pattern of expression of Cyp3a11, whereas the absence of LXR leads to an increase in PB-mediated activation of Cyp2b10. Characteristically, xenobiotic-metabolizing members of the CYP superfamily are transcriptionally induced upon exposure to xenobiotics. This phenomenon increases the ability of the organism to adaptively defend itself against toxic foreign compounds. Since the discovery of CYP induction by the barbiturate phenobarbital (PB) 40 years ago (4), this molecule has served as a prototype for a large group of structurally and functionally diverse compounds that induce the expression of CYP2B genes (5), whereas the glucocorticoid dexamethasone and the antibiotic rifampicin represent drugs that increase CYP3A levels in humans (6). There is, however, considerable overlap between the different inducers and the patterns of genes that they induce. The induction of drug-metabolizing CYP has impor...
tant clinical consequences: 1) it affects the efficiency of drug treatment; 2) it may cause drug-drug interactions; and 3) it can influence endogenous regulatory pathways.

Numerous studies have established that members of the nuclear receptor superfamily of transcription factors are essential in mediating this drug response (reviewed in 7). Specifically, the constitutive androstane receptor (CAR), the pregnane X receptor (PXR), and the chicken xenobiotic receptor have been assigned key roles in the induction of CYP2B, CYP2C, CYP3A, and CYP2H1 genes in human, mouse, and chicken (8–13).

At the same time, CYP-catalyzed reactions are pivotal in the regulation of cholesterol balance and the synthesis and metabolism of steroids and vitamin D. Thus, although cholesterol homeostasis relies on multiple checkpoints, cholesterol 7α-hydroxylase (CYP7A1) is the first and rate-limiting enzyme in the conversion of cholesterol to bile acids (14, 15). Two nuclear receptors, the farnesoid X receptor (FXR) and the liver X receptor (LXR), are important regulators of CYP7A1. Thus, in the mouse, Cyp7a1 is induced by oxysterols via LXR (16, 17). The same gene can be repressed by bile acids via FXR. Here, FXR activates the small heterodimer partner, which in turn inhibits constitutive transcriptional activation by the liver receptor homolog-1 transcription factor on the CYP7A1 promoter (18, 19).

A large body of in vivo and in vitro data support the concept that the molecular mechanisms of hepatic drug induction are linked to endogenous regulatory pathways: observations made in epileptic patients treated with anticonvulsants and in rats treated with PB have revealed increased plasma cholesterol and lipoprotein levels (20–25). Moreover, a study of a 264 bp phenobarbital-responsive enhancer unit (PBRU) from the chicken CYP2H1 gene demonstrated that a DR4-type nuclear receptor response element (direct hexamer repeat separated by four nucleotides) contained within this PBRU was essential for conferring drug induction and that in the LMH chicken hepatoma cell line oxysterols can inhibit the PB induction mediated by this 264 bp PBRU through competitive binding of LXR to the DR4 element (26).

To study the link between LXR, cholesterol homeostasis, and drug induction in vivo, we designed experiments in mice lacking one or both LXR isoforms. Wild-type, LXRα-, LXRβ-, and LXRα/β-deficient mice were kept on standard laboratory chow or a 2% enriched cholesterol diet for 1 week and were given either vehicle (0.9% saline solution) or PB intraperitoneally. Subsequently, levels of Cyp7a1, Cyp2b10, and Cyp3a11, two murine drug-metabolizing enzymes, as well as cholesterol and bile acid transporters were quantified by real-time PCR. Cholesterol and triglyceride (TG) levels also were determined from the animals’ livers.

In this report, we provide evidence for important crosstalk between xenobiotic- and oxysterol-sensing nuclear receptors in the regulation of enzymes involved in drug and cholesterol metabolism. Our data reveal differential regulatory patterns for Cyp2b10 and Cyp3a11 dependent on the expression of LXR isoforms and on challenge of cholesterol metabolism by excess dietary cholesterol. Our results suggest that in the mouse, liver cholesterol status significantly alters the pattern of expression of Cyp3a11, whereas the absence of LXR leads to an increase in PB-mediated activation of Cyp2b10. Finally, we demonstrate that under certain conditions, PB treatment not only affects drug-metabolizing enzymes but also influences hepatic cholesterol levels.

**MATERIALS AND METHODS**

**Reagents**

22(R)-Hydroxycholesterol, 22(S)-hydroxycholesterol, 25-hydroxycholesterol, and pregnenolone 16α-carbonitrile (PCN) were purchased from Sigma (distributed by Fluka AG). 24(S),25-Epoxycholesterol was a kind gift from Thomas A. Spencer (Department of Chemistry, Dartmouth College, Hanover, NH). 27-Hydroxycholesterol was obtained from Steraloids (Newport, RI).

**Plasmids**

The coding regions of mouse CAR and mouse LXRs were amplified from mouse cDNA, followed by subcloning into the pSG5 expression vector (Stratagene, Basel, Switzerland). The expression vector for mouse PXR, pSG5-mPXR, was kindly provided by Steven A. Kliewer (University of Texas Southwestern Medical Center, Dallas, TX).

The pDNA3 expression vector was obtained from Invitrogen AG (Basel, Switzerland). The pDNA3/VP16 construct was provided by Dieter Kressler (Bioszentrum, University of Basel, Switzerland). Vp16/mPXR was generated by PCR amplification of the mouse PXR from pSG5-mPXR and by insertion into the XbaI-digested pDNA3/VP16 vector; the primers harboring XbaI restriction sites were chosen as follows: forward primer, 5’-GCTCTAGAAGGCTGTTACGAGAGGAGCTGAGGCC-3’; reverse primer, 5’-GCTCTAGATCCATGGCTGTCGTGATGAAATAAC-3’. Vp16/mLXRα was generated by amplification of the mouse LXRs from pSG5-mLXRα and by insertion into the digested EcoRI pDNA3/VP16 vector. The primers harboring EcoRI restriction sites were chosen as follows: forward primer, 5’-CTGCTCTAGAAGGCTGTTACGAGAGGAGCTGAGGCC-3’; reverse primer 5’-GGGAAATTCTCACTGGTGACATCCCGAGGCGTTCGCTCT-3’. The pSVβGal expression vector for β-galactosidase used for the normalization of transfection efficiencies was obtained from Promega (Promega Corp., Catalyag AG, Wallisellen, Switzerland).

The pGL3-basic plasmid containing 13 kb of human CYP3A4 5’ flanking region was a gift from Christopher Liddle (University of Sidney at Westmead Hospital, Westmead, Australia). The xenobiotic-responsive enhancer module (XREM; bases 1634 Journal of Lipid Research

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Animals

LXRx-, LXRxβ-, and LXRxβ/β-deficient mice were generated as described previously (29, 30). All LXRx knockout mice used in our study were between 12 and 16 weeks old and had a similar mixed genetic background based on 129/Sv and C57BL/6 strains. Mice with pure 129/Sv background were backcrossed into C57BL/6 mice for seven generations (29, 30). LXRx-deficient mice and C57BL/6 wild-type control mice were bred by Taconic M&B. After delivery, all mice were allowed to adjust for 2 weeks and were housed with a regular 12 h light/12 h dark cycle. Mice were fed ad libitum either a low-fat standard rodent chow diet (R36; Lactamin AB, Vadstena, Sweden) or diet supplemented with 2% (w/w) cholesterol for 1 week. Female mice (n = 5–8) were kept on standard or cholesterol-supplemented chow and were injected one time intraperitoneally with vehicle alone (saline) or PB (100 μM); reverse primer, 5′-GGCCTTTCTGTGTCCTATGCCTTA-3′ (300 nM). Levels of GAPDH were used for normalization. All other primers and probes were chosen as indicated in Table 1.

Transcriptional activation assays

Transactivation assays were carried out in CV-1 green monkey kidney cells as described previously (32). Briefly, cells were expanded for 3 days in DMEM-F12 without phenol red supplemented with 10% charcoal-treated FBS. Subsequently, cells were plated onto 96-well dishes at a density of 25,000 cells/well and grown overnight. For transfection, cells were maintained in Opti-Mem (Invitrogen). Transfection mixes contained 20 ng of receptor expression vector, 8 ng of reporter vector, 60 ng of pSV-βGal, and carrier plasmid to a total of 100 ng of DNA per well. Cells were transiently transfected using the LipofectAMINE reagent (Invitrogen) according to the manufacturer’s protocol. After 24 h of incubation, cells were exposed to drugs or vehicle for an additional 24 h. Transfections with vectors expressing constitutively active nuclear receptors were carried out for 48 h, and then cells were lysed. Cell extracts were prepared using 100 μl of passive lysis buffer (Promega Corp., Catalys AG) and a Wallac 1420 Multilabel Counter. β-Galactosidase activities were measured as described previously (13). Luciferase levels were then normalized against

<table>
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<tr>
<th>Target Gene</th>
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<th>Concentration</th>
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<tr>
<td>Cyp2b10</td>
<td>Forward: 5′-CAATGTTATAGGGAGAAGACTCGG-3′</td>
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<tr>
<td></td>
<td>Reverse: 5′-CAGCTGGAGAGAAGCTCGG-3′</td>
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<td>Probe: 5′-FAM-CCCAAGAAGAGGATGACTACATGCTGATG-3′</td>
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<tr>
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<td></td>
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<td>Probe: 5′-FAM-CGGCCAGAGGAAACCTGGCGCCATGATG-3′</td>
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FAM, 6-carboxyfluorescein; TAMRA, 6-carboxy-tetramethylrhodamine.
β-galactosidase values to compensate for the variation in transfection efficiencies and against untreated control cells to obtain relative luciferase activities.

RESULTS

LXR deficiency and its effect on the basal expression of drug-metabolizing enzymes

To investigate the effects of LXRα, LXRβ, or LXRα/β deficiency on the expression of Cyp2b10 and Cyp3a11, we measured the mRNA levels of these genes in the livers of mice receiving normal diet and treated with vehicle. As illustrated in Fig. 1A, Cyp2b10 and Cyp3a11 show different regulation patterns. Although the absence of one or both LXR isoforms did not significantly affect levels of Cyp2b10 transcript, the basal level of Cyp3a11 mRNA was significantly increased in LXRα/β-deficient mice. Hepatic cholesterol levels were measured in mice on normal diet (Fig. 1B), and in accordance with previously published data (30), total liver cholesterol was found to be increased significantly in LXRα/β-deficient mice.

Effect of dietary cholesterol on the expression of Cyp3a11 and Cyp2b10

Mice with different LXR genotypes were fed with a 2% cholesterol diet for 1 week, after which the expression of Cyp2b10 and Cyp3a11 (Fig. 2) was evaluated. The efficiency of the diet in activating LXR was tested by measuring Cyp7a1 mRNA levels under both normal and high-cholesterol dietary conditions (Fig. 2C). We observed a significant increase in Cyp7a1 expression in wild-type animals when challenged with the 2% cholesterol diet, whereas no significant effect was observed among the other genotypes. Interestingly, 2% cholesterol feeding affected Cyp2b10 and Cyp3a11 expression in a genotype-dependent manner. Expression of Cyp3a11 and Cyp2b10 mRNA is shown in Fig. 2A, B, respectively. In wild-type animals and LXRβ-deficient mice, dietary cholesterol did not significantly alter the expression of either gene. In LXRα- and LXRα/β-deficient mice, however, mRNA levels of Cyp3a11 were increased significantly in response to cholesterol; Cyp2b10 expression levels followed the same profile, but the changes were not statistically significant. This pattern of induction correlates with the liver cholesterol content profiles (Fig. 2D), in which cholesterol is strongly increased in LXRα- and LXRα/β-deficient mice. These results suggest that the influence of cholesterol feeding on Cyp2b10 and Cyp3a11 expression occurs when LXRα is absent and liver cholesterol content is high, partially as a result of the lack of Cyp7a1 induction by LXRα.

Cholesterol feeding reduces PB-mediated increase of Cyp2b10 independently of LXR genotype

As shown previously in vivo (33), PB massively induces Cyp2b10 (140-fold) in wild-type mice under standard dietary conditions (Fig. 3A). In mice lacking LXRβ or both LXR isoforms, this strong PB-mediated induction was increased further (Fig. 3A), suggesting that the absence of LXR positively influences the capacity of Cyp2b10 to be induced by PB. When wild-type or LXR-deficient mice are treated with PB while fed a 2% cholesterol diet, decreased inducibility by PB was observed, irrespective of LXR genotype (Fig. 3A). This repression of PB induction by dietary cholesterol thus seems to be LXR-independent, which implies that the hepatic cholesterol status affects the regulation of the drug-metabolizing enzyme Cyp2b10.

Effect of cholesterol feeding on PB-mediated induction of Cyp3a11

To study the influence of dietary cholesterol on the PB-mediated increase of Cyp3a11, mRNA levels were determined in mice fed standard chow or a 2% cholesterol diet and treated with either vehicle or PB (Fig. 3B). In mice of all LXR genotypes, PB induced Cyp3a11 expression. However, this PB-mediated increase was smaller in LXRα/β-deficient mice, whereas the basal level of Cyp3a11 expression was higher than in the wild type (Fig. 1A). In contrast to Cyp2b10, cholesterol feeding did not significantly reduce PB-mediated induction of Cyp3a11 mRNA in any of the mouse genotypes. PB induction of Cyp3a11 is thus apparently independent of dietary cholesterol content.

PB treatment affects hepatic TG and cholesterol levels

To evaluate the effect of PB on cholesterol homeostasis and lipogenesis, hepatic cholesterol and TG levels were
measured in mice fed standard laboratory chow or the 2% cholesterol diet (Fig. 4). Compared with wild-type animals, LXRα/β-deficient mice displayed significantly decreased levels of hepatic TG (Fig. 4A) in the absence of drug treatment, consistent with previous observations (30). When mice were fed standard chow, PB treatment had no significant effect on hepatic TG. In contrast, PB significantly decreased hepatic TG levels in wild-type, LXRα-deficient, and LXRβ-deficient animals but not in LXRα/β-deficient mice (Fig. 4B) on the 2% cholesterol diet. Furthermore, hepatic cholesterol levels of LXRα/β-deficient mice were increased significantly compared with the other genotypes when mice were fed standard chow (Fig. 4C), in agreement with previously published data (30). Most interestingly, PB treatment was found to decrease hepatic cholesterol levels in mice on the high-cholesterol diet, and this
Effect was independent of their LXR genotype (Fig. 4D). In mice kept on the normal diet, PB did not further decrease hepatic cholesterol levels, with the exception of LXRβ-deficient (−/−) mice (Fig. 4D), which exhibited increased cholesterol levels already under normal dietary conditions (Fig. 4C). The occurrence of this effect in LXRβ-deficient mice suggests that the capacity of PB to decrease hepatic cholesterol levels is independent of LXR. Interestingly, the effect of PB occurs only when hepatic cholesterol and TG levels are increased.

**PB affects hepatic cholesterol levels by acting on Cyp7a1 and MRP3 expression**

To gain insight into the pathways by which PB modifies hepatic cholesterol levels, Cyp7a1 mRNA levels were quantified by Taqman analysis (Fig. 5A). In addition to the increase observed in wild-type vehicle-treated mice upon cholesterol feeding, a PB-mediated increase in Cyp7a1 expression was measured in LXRβ-deficient mice under both feeding conditions. PB treatment did not lead to significant changes in Cyp7a1 expression in the other genotypes (data not shown), suggesting that this effect occurs predominantly in the total absence of LXR.

To examine the molecular details underlying the activation of Cyp7a1, we cloned a tandem repeat of a previously identified LXR response element from the Cyp7a1 gene (27) into a luciferase reporter vector under the control of the thymidine kinase promoter. This reporter construct, (DR4)2tkluc, was then used in transcriptional activation assays in CV-1 cells (Fig. 5B, C). As expected, reporter activity was increased when cells were cotransfected with a VP16mLXRα construct expressing a constitutively active derivative of the mouse LXRα isoform (Fig. 5B). It is well established that PB mediates the activation of drug-metabolizing CYP3A and CYP2B enzymes via PXR and CAR (5). For this reason, we tested whether these two nuclear receptors interact with the known regulatory sequence in the mouse Cyp7a1 gene in vitro. Interestingly, (DR4)2tkluc reporter activity was strongly increased when cells were cotransfected with a plasmid expressing a constitutively active mouse PXR-VP16 fusion protein (Fig. 5B).

When cells were cotransfected with an expression vector for mouse CAR (Fig. 5C), a 3-fold increase of basal activation was observed. The activity of mouse CAR can be modulated by androstanol, an inverse agonist, and by the inducer compound TCPOBOP [1,4-bis(3,5-dichloropyridyloxy) benzene], as has been reported (34). Using these compounds, we could modulate the expression of the (DR4)2tkluc reporter (Fig. 5C) to a comparable extent as reported for bona fide CAR target genes. These data implicate PXR and CAR in the regulation of mouse Cyp7a1, particularly in the absence of LXR's, and suggest that the cholesterol-lowering effect observed in the pres-

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**Fig. 4.** PB treatment affects triglyceride (TG) and cholesterol levels. Wild-type (WT), LXRα−, LXRβ−, and LXRα/β-deficient (−/−) mice (n = 5–8), fed standard laboratory chow or a 2% cholesterol-supplemented diet for 1 week, were administered either vehicle (saline) or PB (100 mg/kg) for 16 h. TG contents in livers from animals fed standard chow (A) or 2% cholesterol (B) as well as total cholesterol contents in animals fed standard chow (C) or 2% cholesterol (D) and treated with vehicle or PB were determined. Error bars represent SEM. * P < 0.05 determined by Student’s t-test.
Cholesterol derivatives are activators of mouse PXR

To assess the potential of cholesterol and several of its derivatives to activate mouse PXR, transcriptional activation assays were performed in CV-1 cells (Fig. 6). PCN, a known potent activator of murine PXR used as a positive control, led to 18-fold activation of the reporter gene activity over vehicle-treated cells. In addition, hydroxylated derivatives of cholesterol, like the potent LXR agonist 22(S,25)-epoxycholesterol, also could activate PXR, confirming previous observations in mouse hepatocytes (39). 22(R)-Hydroxycholesterol displayed only weak activation of PXR, whereas 22(S)-hydroxycholesterol was inactive. Interestingly, however, MRP3 expression was increased in all genotypes by PB when mice were fed cholesterol (Fig. 5D), suggesting that bile salt export from the liver is increased in those animals in which liver cholesterol was increased. PB treatment also increased MRP3 expression in wild-type mouse liver, confirming previous observations (38).

**DISCUSSION**

In this report, we describe the influence of LXRα and/or LXRβ deficiency and the effect of cholesterol feeding on the expression and PB-mediated induction of two murine drug-metabolizing enzymes, Cyp2b10 and Cyp3a11. In addition, we describe the consequences of a short-term PB treatment on hepatic cholesterol and TG content in mouse livers.

Our data clearly show that LXR deficiency differentially affects not only the basal expression levels of Cyp2b10 and Cyp3a11 but also the capacity of these genes to be induced by PB. Upon exposure to dietary cholesterol, Cyp2b10 and Cyp3a11 exhibited a gene-specific response. Although no significant effect of LXR deficiency was observed on the

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**Fig. 5.** PB treatment affects the hepatic expression of Cyp7a1 and multidrug resistance-associated protein 3 (MRP3). A: Wild-type and LXRα/β-deficient mice (n = 5–8), fed standard laboratory chow or a 2% cholesterol-supplemented diet for 1 week, were administered either vehicle (saline) or PB (100 mg/kg) for 16 h. Hepatic Cyp7a1 expression was determined by real-time PCR. Error bars represent SD. * P < 0.05 determined by Student's t-test. B: A luciferase reporter vector containing two copies of the mouse Cyp7a1 LXR response element, a direct-repeat separated by four nucleotides, (DR4)2tkluc, was cotransfected in CV-1 cells together with expression vectors for VP16mLXRα, VP16mPXR, or a control vector. Cell extracts were analyzed for luciferase activities, which then were normalized against β-galactosidase activities. Luciferase activities relative to cells cotransfected with empty vector are shown. Values represent means of at least three independent experiments ± SD. C: (DR4)2tkluc was transfected in CV-1 cells with or without an expression vector for mouse constitutive androstane receptor (mCAR). Cells were treated for 24 h with vehicle (0.1% DMSO), androstanol (And; 5 μM), and/or TCPOBOP [1,4-bis(3,5-dichloropyridyloxy)benzene] (10 μM). Cell extracts were analyzed as described above. Absolute luciferase activities are shown. Values represent means of at least three independent experiments ± SD. D: MRP3 levels were determined by real-time PCR in the indicated mouse livers. Gene levels are expressed relative to the wild-type (WT) vehicle-treated control. Error bars represent SD. * P < 0.05 determined by Student's t-test.
expression of Cyp2b10, Cyp3a11 was increased significantly in mice lacking both LXR isofoms. This increase may be either a direct consequence of the lack of LXRs and LXRβ (derepression of gene activation) or a response to secondary events occurring in mouse liver linked to LXR deficiency. Therefore, we determined hepatic cholesterol levels and found them to be significantly higher in LXRs/β-deficient mice compared with wild-type animals. Cyp3a11 expression was increased in mice fed cholesterol and displaying higher hepatic cholesterol content, whereas no increase was seen in Cyp2b10 expression. Studies in vitro have shown that CYP3A4 can metabolize cholesterol to 4β-hydroxycholesterol and that this metabolite is present at relatively high concentrations in human plasma (40). This metabolic conversion was shown to occur only for CYP3A4 and not for CYP1A1, CYP2C9, or CYP2B6. Although it is currently unknown whether the mouse homolog Cyp3a11 metabolizes cholesterol like CYP3A4, a possible interpretation of our findings is that cholesterol could act as a likely substrate for Cyp3a11 and may increase its own metabolism when present in the liver in large amounts. Indeed, in wild-type mice, cholesterol feeding leads to an activation of LXR and its target gene Cyp7a1, which leads to an increase in cholesterol metabolism, decreases cholesterol levels, and presumably does not increase Cyp3a11. By contrast, in LXR-deficient mice, cholesterol levels are not controlled by LXR and Cyp7a1; therefore, they are excessive and may induce Cyp3a11.

As murine PXR is well known as a key regulator of Cyp3a11 expression both in vitro and in vivo (41), we suspect that the induction observed can be at least partially attributable to the activation of this nuclear receptor. In addition to cholesterol, the levels of several of its hydroxylated derivatives are increased in mice when one or both LXRs are ablated or when mice are fed cholesterol (42). Thus, it is known that 24(S),25-epoxycholesterol accumulates in the liver after cholesterol feeding (43). Moreover, wild-type mice and mice lacking LXRs/β have been described to have increased circulating levels of 27-hydroxycholesterol when fed 2% cholesterol (29). Rats, kept on a high-fat diet, show a considerable increase in 25-hydroxycholesterol in liver homogenates, whereas 27-hydroxycholesterol levels remain unaffected (44). Based on these observations, and considering the remarkable correlation between hepatic cholesterol levels and Cyp3a11 expression, we evaluated the potential of several hydroxylated derivatives of cholesterol to activate mouse PXR in cell culture. 24(S),25-Epoxycholesterol was a good activator of mouse PXR in our system, in accordance with previous observations in mouse hepatocytes (39). Interestingly, 25- and 27-hydroxycholesterol efficiently activated murine PXR as well. These data suggest that in mice, not only bile acid precursors (45) and bile acids (41) but also other endogenous cholesterol derivatives can regulate Cyp3a11 expression via PXR.

Induction of Cyp2b10 and Cyp3a11 by PB was studied in LXR-deficient and cholesterol-fed mice. Compared with wild-type animals, LXRs- or LXRα/β-deficient mice showed significantly higher induction of Cyp2b10 mRNA levels, suggesting that LXRs can repress PB-mediated activation of Cyp2b10 in vivo in mice. Moreover, when mice were fed cholesterol, the PB-mediated increase in expression was reduced independently of the genotype, suggesting that this dietary effect on induction is not LXR-dependent. For Cyp3a11, the situation is probably different, because whereas dietary cholesterol had no effect on PB induction in wild-type and single-LXR-deficient mice, a significant decrease of induction occurred in LXRs/β-deficient mice. However, the reduced capacity of Cyp3a11 to respond to PB may be a consequence of the high basal expression of the gene in the absence of LXRs, preventing further response. Moreover, when mice were fed cholesterol, no significant difference was observed in the PB-mediated response of Cyp3a11 compared with standard chow feeding, in contrast to our observations with Cyp2b10.

In addition to the effect of PB on murine drug-metabolizing enzymes, we were also interested in studying the effect of short-term PB treatment on hepatic TG and cholesterol levels, especially given that cholesterol and several of its derivatives can act as important signaling molecules. Hepatic TG production is determined mainly by the fatty acid synthesis rate, which is controlled to a large extent at the transcriptional level by peroxisome proliferator-activated receptor α, which stimulates fatty acid β-oxidation (46), and by sterol-regulatory element binding protein 1c (SREBP-1c) (47), which controls fatty acid synthesis. In addition to cholesterol transport genes, LXR has been identified as an important regulator of SREBP-1c and de novo lipogenesis (48). The TG-lowering effect of PB occurred.
only when wild-type, LXRα-deficient, or LXRβ-deficient mice were fed cholesterol and had increased TG before PB treatment. Our data show that PB treatment significantly decreases hepatic cholesterol levels, but only when they are high. The main pathway for the elimination of cholesterol is its conversion to bile acids, which is dependent on Cyp7a1. In mice, positive regulation of Cyp7a1 is mediated by LXRα (17), whereas negative feedback is exerted by bile acids via FXR and the small heterodimer partner (19). Moreover, it has been shown that PCN treatment represses Cyp7a1 expression in vivo in mice and that this effect is PXR-dependent (49). Interestingly, we observed that, in LXRα/β-deficient mice, PB acts as a positive regulator of Cyp7a1 under both dietary conditions, thus potentially explaining the observed decrease in cholesterol levels under these circumstances. A similar observation was made in earlier studies in rats, in which PB treatment led to a decreased content of cholesterol in liver microsomes in a strain that responded with increased Cyp7a1 activity (50). In addition, we showed in vitro that not only PXR but also CAR can interact with the DR4 element present in the 5′ flanking region of the murine Cyp7a1, providing a mechanism for this particular effect of PB. An additional mechanism to reduce cholesterol levels by PB could be the activation of MRP3 and the consequent increased excretion of bile salts.

In conclusion, our data demonstrate that basal and PB-induced expression of Cyp2b10 and Cyp3a11 is differentially and specifically influenced by the absence of LXR as well as the status of cholesterol in the liver. These observations suggest that cholesterol and its hydroxylated derivatives are endogenous signaling molecules, normally serving as ligands to LXR, but under extraordinary conditions, such as stringent diet or the absence of functional LXR, they may also serve as ligands for other nuclear receptors such as PXR or CAR and therefore affect the expression of drug-metabolizing enzymes. Indeed, recent gene expression data in cholesterol-fed mice support the concept that cholesterol or its products increase a number of PXR/ CAR target genes (A. Roth, R. Looser, and U. A. Meyer, unpublished observation). The intricate nuclear receptor signaling network allows a single gene to be a target for several nuclear receptors, sensing different markers of endogenous or exogenous origin. Our findings suggest that processes that influence hepatic cholesterol levels, such as defects in cholesterol biosynthesis pathways or hepatic accumulation of lipids, can ultimately alter drug metabolism, affecting the efficiency of drug treatment.

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