Acyl-CoA:Cholesterol Acyltransferase Promotes Oxidized LDL/Oxysterol-Induced Apoptosis in Macrophages

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Abbreviations: 25-OHC, 25-hydroxycholesterol; 7KC, 7-ketocholesterol; 7KC esters, 7-ketocholesteryl esters; AA, arachidonic acid; AACOCF₃, arachidonyl trifluoromethyl ketone; cPLA₂, cytosolic phospholipase A₂; ETYA, 5,8,11,14-Eicosatetraynoic acid; MPM, mouse peritoneal macrophages; oxLDL, oxidized low-density lipoproteins.
ABSTRACT

7-ketocholesterol (7KC) is a cytotoxic component of oxidized low-density lipoproteins (oxLDL) and induces apoptosis in macrophages by a mechanism involving activation of cytosolic phospholipase A2 (cPLA2). In the current study, we examined the role of acyl-CoA: cholesterol acyltransferase (ACAT) in 7KC-induced and oxLDL-induced apoptosis in murine macrophages. An ACAT inhibitor, Sandoz 58-035, suppressed 7KC-induced apoptosis in P388D1 cells and both 7KC-induced and oxLDL-induced apoptosis in mouse peritoneal macrophages (MPMs). Furthermore, in comparison to wild type MPMs, ACAT-1 deficient MPMs demonstrated significant resistance to both 7KC-induced and oxLDL-induced apoptosis. Macrophages treated with 7KC accumulated ACAT-derived $^{14}$C]cholesteryl and $^{3}$H]7-ketocholesteryl esters. Tandem LC/MS revealed that the 7KC esters contained primarily saturated and monounsaturated fatty acids. An inhibitor of cPLA2, arachidonyl trifluoromethyl ketone, prevented the accumulation of 7KC esters and inhibited 7KC-induced apoptosis in P388D1 cells. The decrease in 7KC ester accumulation produced by inhibition of cPLA2 was reversed by supplementing with either oleic or arachidonic acid; however, only arachidonic acid supplementation restored induction of apoptosis by 7KC. These results suggest that 7KC not only initiates the apoptosis pathway by activating cPLA2, as we have previously reported, but also participates in the downstream signaling pathway when esterified by ACAT to form 7KC-arachidonate.

Supplementary Key Words: 7-ketocholesterol, oxysterol esterification.
The presence of apoptotic macrophages within atherosclerotic lesions of both animals and humans has been consistently observed over the past decade; however, the physiological significance of macrophage apoptosis in atherogenesis is not fully understood. Macrophage apoptosis has been proposed to be a mechanism by which lipids accumulate within the coronary vasculature and thereby contributes to plaque formation and progression (1,2). Alternatively, macrophage apoptosis has been speculated to be as a defense mechanism, perhaps to limit the number of lesional macrophages and the hazardous consequences associated with necrotic cell death within lesions (3). Recent results from our laboratory demonstrated that macrophage apoptosis via the mitochondrial apoptosis pathway has an overall protective role in atherogenesis (4). In this study, mice with genetically altered macrophages deficient for Bax, a pro-apoptotic gene, developed larger and more advanced lesions with reduced apoptosis compared to control mice with normal macrophages. This result underscores the necessity to fully elucidate the mechanisms that regulate macrophage apoptosis and potentially influence the development and progression of atherosclerosis.

Many of the pathological events associated with the development of atherosclerosis are related to the uptake and metabolism of modified low-density lipoproteins (LDL), such as oxidized LDL (oxLDL). The continuous uptake of oxLDL by macrophages in the vascular intima via specific receptor mediated mechanisms and by phagocytosis (5,6,7) results in the formation of lipid-laden “foam cells” that are the defining characteristic of early atherosclerotic lesions (8). The ingestion of oxLDL by macrophages has been shown to be eventually cytotoxic (9,10,11,12) and this cytotoxicity proceeds, at least in part, through induction of the mitochondrial apoptosis pathway (13,14).
Cholesterol oxidation products found in oxLDL, collectively referred to as oxysterols (15,16), have been demonstrated to largely account for the apoptotic activity of oxLDL in macrophages (17,18). Oxysterols are commonly found in foods of animal origin (19) and have been demonstrated to exist both in oxLDL (20) and in atherosclerotic plaque (21). The predominant oxysterol present in oxLDL is 7-ketocholesterol (7KC), accounting for up to 30% of the total sterols in extensively oxidized LDL (22). 7KC in the 10-25 µM range induces apoptosis in macrophages, vascular endothelial cells, and smooth muscle cells (23). 25-hydroxycholesterol (25-OHC) is another oxysterol found in oxLDL, albeit at much lower concentrations than 7KC (22). 25-OHC has been used extensively as a model oxysterol compound in studies of both oxysterol toxicity as well as cellular cholesterol homeostasis. 25-OHC induces apoptosis in cultured monocyte-macrophages (13,24,25), and lymphoid cell lines (26,27), in the range of 1-10 µM. The concentrations of oxysterols used to induce apoptosis in these in vitro studies are below the upper limit of the physiological concentration of oxysterols detected in human plasma following the ingestion of a test meal, which has been reported to be as high as 37 µM (28) and with in the range of oxysterols most commonly detected in plasma of individuals with hypercholesterolemia, 10-30 µM (29,30,31). Like oxLDL, oxysterols induce the release of cytochrome c from mitochondria (32,33), the hallmark of the mitochondrial apoptosis pathway.

Our previous work demonstrated that the mechanism by which oxysterols (25-OHC and 7KC) induce apoptosis involves an increase in cytosolic calcium and subsequent activation of cytosolic phospholipase A2 (cPLA2), and that loss of cPLA2 activity, either by use of specific inhibitors or by cPLA2 gene knockout, results in resistance to oxysterol-induced apoptosis in macrophages (13,34). We have also previously shown that macrophages treated with
5,8,11,14-eicosatetraynoic acid (ETYA), an inhibitor of arachidonic acid (AA) metabolism, are resistant to the induction of apoptosis by 25-OHC or 7KC (13,34) indicating that metabolism of cPLA₂-released AA may be required for the induction of apoptosis by these oxysterols.

OxLDL also activates cPLA₂ in macrophages and the fatty acids that are released as a result of this activation have been shown to become esterified to oxLDL-derived cholesterol (35). Intracellular cholesterol esterification is catalyzed by the microsomal enzyme acyl-coenzyme A:cholesterol acyltransferase (ACAT), which uses cholesterol and fatty acyl coenzyme A as substrates (36, 37). In addition to cholesterol, a variety of oxysterols are also ACAT substrates (38) and the oxysterols found in atherosclerotic plaques are predominantly esterified (39). This suggests that the fatty acids released by oxysterol-induced activation of cPLA₂ in macrophages may be used as substrates in esterification reactions catalyzed by ACAT and that ACAT potentially plays a role in the apoptotic signaling pathway induced by oxysterol treatment of macrophages.

Therefore, we undertook the current study to examine the hypothesis that ACAT plays a role in oxLDL/oxysterol-induced apoptosis in murine macrophages. For this study, we utilized the well-studied P388D1 macrophage cell line as a model and then verified our findings in isolated murine primary macrophages (MPMs) as we have previously done to further elucidate the oxysterol apoptotic signaling mechanism in macrophages (13,34). Our results reveal that loss of ACAT activity in macrophages, either by pharmacological inhibition or by genetic knockout of ACAT-1, results in reduced induction of apoptosis in response to 7KC or oxLDL. Furthermore, we provide evidence that the apoptotic signal generated in macrophages by ACAT may be an arachidonyl oxysterol.
MATERIALS AND METHODS

Materials

P388D1 were purchased from American Tissue Type Collection (Manassas, VA). BioWhittaker cell culture media and Falcon tissue culture plates were purchased from Fisher Scientific. NovaCell 1 fetal bovine serum (FBS) was purchased from Nova-tech, Inc (Grand Island, NE). All other tissue culture supplements were purchased from InVitrogen (Carlsbad, CA). The acyl-CoA:cholesterol acyltransferase inhibitor, 3-[Decyldimethylsilyl]-N-[2-(4-methylphenyl)-1-phenylethyl]propanamide (58-035), was kindly provided by Novartis Pharmaceuticals Corp. (formerly Sandoz, Inc.) (East Hanover, NJ). The following were obtained from BIOMOL Research Laboratories Inc. (Plymouth Meeting, PA); Arachidonyl trifluoromethyl ketone (AACOCF₃), 5,8,11,14-Eicosatetraynoic acid (ETYA), 3β-(2-diyethylaminoethoxy)-androstenone (U-18666A), caspase-3 assay substrate (Ac-DEVD-AFC) and inhibitor (Ac-DEVD-CHO). 7-ketocholesterol was obtained from STERALOIDS, Inc. (Newport, RI). All radiolabeled chemicals, [1,2,6-³H]7-ketocholesterol, [¹⁴C]cholesterol and [1-¹⁴C]arachidonic acid were purchased from American Radiolabeled Chemicals, Inc. (St. Louis, MO). Oleoyl chloride, arachidonoyl chloride, stearoyl chloride, linoleoyl chloride, and palmitoyl chloride were purchased from Sigma-Aldrich Chemical Company (St. Louis, MO). Silicic acid (100-200 mesh) was purchased from Bio-RAD laboratories (Richmond, CA). Organic solvents and reagents were of analytical or HPLC grade and were obtained from either Fisher Scientific (Pittsburgh, PA) or Sigma-Aldrich Chemical Company (St. Louis, MO.).
Mice

C57Bl/6J mice were purchased from Jackson Laboratory (Bar Harbor, ME). ACAT-1 deficient mice were bred and fully backcrossed to C57BL/6J genetic strain at Vanderbilt University School of Medicine to allow direct comparison to wild type C57Bl/6J mice and have been described previously (40). All animal procedures described in this study were performed in accordance with guidelines administered by the Animal Research Facility at East Tennessee State University Quillen College of Medicine or by the Institutional Animal Care and Usage Committee at Vanderbilt University School of Medicine.

Cell culture

All cells were maintained in a humidified atmosphere of 5% CO_2 and 95% air at 37°C as previously described (34). P388D1 cells were cultured in Dulbecco’s modified Eagle’s medium supplemented with 10% fetal bovine serum, 2 mM glutamine, 1 mM sodium pyruvate, 100 units/ml penicillin, and 100 µg/ml streptomycin.

Preparation of oxLDL

LDL (1.019 < d < 1.063) was prepared from normal human serum by sequential ultracentrifugation, extensively oxidized by incubation with CuCl_2, and evaluated as previously described (11). The oxLDL was extensively dialyzed against PBS containing 200 µM EDTA at 4°C, stored in the dark and used within two weeks.

Caspase-3 assay

Log phase P388D1 cells were seeded at 2x10^6/well in 12-well culture plates in 1 ml of cell culture media (DMEM + 10%FBS). One hour prior to the addition of 7KC or oxLDL, inhibitors were diluted directly into the media from stock solutions made in DMSO while controls received an equivalent volume of vehicle. 7KC (from 2 mg/ml ethanol stock solutions)
or oxLDL were then added to the wells while controls received an equivalent volume of vehicle. The cells were incubated for 16 h, harvested, and assayed for caspase-3 and caspase-3 like activity using a fluorogenic substrate (Ac-DEVD-AFC) and a spectrofluorometer equipped with a microplate reader as previously described (34). The relative fluorescent units were normalized to the protein concentration of the sample, which was determined by µBCA assay (Pierce, Rockford, IL).

**Analysis of apoptosis in mouse peritoneal macrophages**

Mouse primary macrophages (MPMs) were isolated by lavage of the peritoneal cavity and cultured essentially as described previously (13). Isolated MPMs were plated at a density of 1x10^5/ well in 8 well glass chamber slides (Nalge Nunc International, Naperville, IL) in DMEM+10% FBS and allowed to attach for 24 h in a humidified 5% CO₂ incubator at 37 °C. Unattached cells were removed by washing twice with PBS. The MPMs were preincubated in DMEM+10% FBS supplemented with and without inhibitors for 1 h prior to the addition of 7KC or oxLDL as indicated in the figure legends. Control wells received an equivalent volume of vehicle. After incubating for 24 h or 48 h, the cells were rinsed with PBS and fixed in 4% buffered paraformaldehyde for 30 min at room temperature. The cells were then rinsed with PBS, permeabilized, and subjected to DNA end labeling with isothiocyanate-dUTP utilizing a commercially available *in situ* apoptosis detection kit as previously described (13). The TUNEL assay was performed blind and the TUNEL-positive cells were identified by observation using a Nikon Diaphot-200. For each treatment, the percent of TUNEL-positive cells in ten randomly chosen fields (45 to 60 cells/field) was determined.
Determination of cholesteryl and oxysterol ester formation

P388D1 cells were seeded as described above for caspase-3 assays. The culture medium was supplemented with and without inhibitors while the controls received an equivalent volume of vehicle. Following a 1 h incubation, tracer amounts of a radiolabeled oxysterol (1 µCi/ml of [3H]7-ketocholesterol) or cholesterol (0.5 µCi/ml of [14C]cholesterol) was added. At this time, unlabeled oxysterols were added (controls received an equivalent volume of vehicle) and the incubation continued for 16 h. The cells were collected, rinsed with ice-cold PBS twice, and the lipids were extracted with hexane:isopropanol 60:40 (v/v). The lipids were dried under a stream of N₂ gas, resuspended in hexane, and subjected to thin layer chromatography (TLC) on silica gel 60 plates with hexane:diethyl ether:acetic acid 80:20:1 (v/v/v) as the mobile phase. Using this system, esters of 7KC and cholesterol migrate farther (Rf~0.41 and Rf~0.7, respectively) than unesterified 7KC and cholesterol, which remain at the origin. The plates were dried, saturated with En³Hance (PerkinElmer Life And Analytical Sciences, Inc., Boston, MA), and subjected to autofluorography. The identity of ACAT derived products was inferred by their inhibition with 58-035 and when possible by their co-migration with authentic standards. The areas corresponding to 7KC or cholesteryl esters were scraped from the plate and the radioactivity measured by liquid scintillation counting (LSC). The lipid extracted cell pellets were dissolved in 0.1 N NaOH and the protein concentrations were determined by μBCA assay (Pierce, Rockford, IL). All treatments were done in triplicate and the data is presented as mean dpm/mg protein ± S.D.

Analysis of 7-ketocholesteryl esters by tandem LC/MS

Log phase P388D1 cells (2.0 x 10⁶/ml) were cultured in DMEM+10%FBS supplemented with 7-ketocholesterol (20 µg/ml) for 16 h at 37°C. The cells were then collected, rinsed with
ice-cold PBS twice, and the intracellular lipids extracted with hexane:isopropanol 60:40 (v/v) in
the presence of butylated hydroxyl toluene (50 µg/ml) at room temperature for 20 min. The
mixture was centrifuged for 15 min at 1,000 x g and the lipid extracts were placed in a clean
glass vial, dried under a stream of N₂ gas, resuspended in chloroform:methanol (3:1) and
stored at -80°C until analyzed.

Similar to Jessup and colleagues (22), we synthesized five 7-ketocholesteryl esters:
palmitate, stearate, oleate, linoleate, and arachidonate. The synthesis of these 7-
ketocholesteryl esters was essentially by means of modification of the method of Piretti and
Pagliuca (41). Equal molar amounts of 7KC and fatty acyl chlorides were dissolved in 0.5 ml
of anhydrous pyridine and incubated at 56°C for three hours. The reaction products were
extracted into petroleum ether and the organic phase was placed into a clean glass tube and
dried under nitrogen. After evaporating the solvent, the crude residue was resuspended in
hexane and the free 7KC was separated from the esterified 7KC on silica gel columns (42).
The purity of the 7KC esters was greater than 98% as determined by reverse phase high
performance liquid chromatography on a Versapack C18 column (Alltech, Nicholasville, KY),
using a mobile phase of acetonitrile-isopropanol-water (44:54:2 v/v/v) and detected at 234 nm
essentially as described (22). The purified 7KC esters were dissolved in chloroform:methanol
3:1 (v/v) and stored at -80°C. The concentration of the 7KC esters was determined using an
Amplex Red Cholesterol Assay Kit (Molecular Probes, Eugene, OR) with 7-ketocholesterol as
the standard.

Equimolar mixtures of the five 7KC ester (16:0, 18:0, 18:1, 18:2 and 20:4) standards
were prepared in isopropanol to give a concentration of 0.1, 1.0 and 10.0 µM. The
pentafluorophenyhydrazone (PFPH) derivatives were prepared by treatment of a 100 µL
aliquot of each standard with 100 µL of 10 mM pentafluorophenylhydrazine in isopropanol, followed by addition of 20 µL glacial acetic acid. The reaction mixture was heated at 60 °C for 1 hour and then analyzed without further dilution by LC/MS. A 100 µL aliquot (from 1 mL total) of an extract from macrophage lipids was derivatized by this method.

The 7KC ester PFPH derivatives were analyzed by tandem LC/MS using a ThermoElectron Corp. (San Jose, CA) Quantum Discovery triple quadrupole LC/MS instrument equipped with a Surveyor HPLC and autosampler. Samples were ionized using the standard electrospray ion (ESI) source. The ESI source was operated at 4.0 kV, using nitrogen for both the nebulizer sheath gas and auxiliary drying gas. The ion transfer capillary was operated at 325 °C with a source declustering potential of 10 volts. The collision cell was operated at an argon pressure of 1.5 mTorr. Survey studies in tandem MS mode showed that the most prominent product ion was derived from scission of the fatty acyl-sterol bond to give an ion at m/z 563 due to loss of the fatty acid as a neutral species. The optimum collision energy was 32 volts for all of the 7KC esters that were examined. Quantitation was performed by selected reaction monitoring (SRM) using the [M + H] precursor ions for selected C16, C18 and C20 esters and the common product ion at m/z 563. LC/MS separations were performed on a 2.1 x 150 mm Zorbax XDB-C8 5 µ silica column using isocratic elution at 300 µL/min. The elution solvent was acetonitrile:isopropanol:water (50:45:5) containing 0.5% acetic acid.

Quantitation was done by the method of external normalization of individual 7KC esters using accurately prepared complex mixtures of the five standards. Thus, 20 µL of a 0.1 µM standard (2 pmol total) was used to establish the absolute LC/MS response factor (ARF) for individual standards in area counts/pmol. The ARF was used to calculate the absolute amount of individual lipid species injected into the instrument in pmol based on accurate
extraction and dilution of total lipids in macrophages. The relative response factors (RRF) for individual 7KC esters ranged from 0.14 (18:1) to 1.0 (18:0); these were used to calculate the normalized amount of individual species present in a macrophage lipid extract. Cell culture samples were analyzed in triplicate and the means reported. The absolute detection limit by this method is approximately 10 – 50 fmoles, depending on the fatty acyl species.

**Statistical analysis**

Data are presented as mean ± standard deviation of triplicate treatments. Student's t-test was used to analyze the significance of differences between independent samples. \( P < 0.05 \) was considered as statistical significance.
RESULTS

Pharmacological inhibition of ACAT prevents 7KC-induced apoptosis in a murine macrophage cell line.

To determine if ACAT activity has a role in the induction of apoptosis by 7KC in macrophages we tested the ability of a well-characterized ACAT inhibitor, Sandoz compound 58-035 (43), to effect 7KC-induced apoptosis in P388D1 cells. Control experiments were performed (data not shown) in which the incorporation of labeled oleate into cholesterol ester was used to determine dosages of 58-035 which could produce the loss of detectable cholesterol esterification in macrophage cell lines and mouse peritoneal macrophages that has been reported by others (44). Treatment of P388D1 cells with 58-035 (10 µg/ml) effectively blocked the induction of caspase-3 activity by 7KC (Fig. 1). 58-035 also prevented the induction of apoptosis by 7KC in Raw 264.7 macrophages (data not shown). A similar result was observed when P388D1 or Raw 264.7 cells were incubated with 25-OHC in the presence of 58-035 (data not shown); however, 58-035 had no effect on the induction of apoptosis by staurosporine, a protein kinase inhibitor well known to induce apoptosis via the mitochondrial pathway (data not shown).

An ACAT inhibitor attenuates 7KC and oxLDL-induced apoptosis in isolated resident peritoneal macrophages.

We also analyzed the effects of 58-035 on apoptosis induced by 7KC in isolated MPMs. Isolation of resident MPMs does not yield enough cells to accurately quantitate caspase-3 activity by our method unless we pool MPMs isolated from a large cohort of mice. Therefore, we utilized an in situ terminal transferase-mediated dUTP nick end-labeling (TUNEL) method.
to quantitate the apoptosis. The TUNEL assay has been demonstrated to be a highly sensitive and specific method for detecting individual apoptotic cells grown on cover slips (45) and has been previously employed by our laboratory to quantitate apoptosis in MPMs (13). Treatment of MPMs with 7KC (10 µg/ml) for 24 h induced an ~5.8 fold increase in the percentage of TUNEL positive cells compared to untreated controls (Fig. 2A). When the medium was supplemented with 58-035, the induction of TUNEL positive MPMs by 7KC was prevented (Fig. 2A). Apoptosis was also induced in MPMs by treatment with 25-OHC, which was effectively prevented by 58-035 (data know shown).

Incubation with 50 µg/ml oxLDL for 24 to 48 h is commonly utilized to induce macrophage foam cell formation in vitro (35). In addition, the degree of apoptotic induction we observed using 10 µg/ml 7KC (Fig. 2A), is nearly identical to the degree of apoptotic induction we previously reported when MPMs were incubated with 50 µg/ml oxLDL for 24 h (13). We therefore performed TUNEL analysis of MPMs that had been incubated with 50 µg/ml oxLDL for 24 and 48 h in the presence and absence of 58-035. Treatment with oxLDL produced an ~3.0 fold increase in TUNEL positive cells after 24 h and an ~4.0 fold increase after 48 h (Fig. 2B). In the presence of the ACAT inhibitor, the induction of TUNEL positive cells by oxLDL was reduced to only ~1.7 fold over controls at 24 h and ~2.0 fold over controls at 48 h.

**ACAT-1 deficient macrophages display significantly reduced levels of apoptosis in response to 7KC or oxLDL.**

ACAT-1 is the primary ACAT isoform expressed by macrophages and peritoneal macrophages isolated from ACAT-1 (-/-) mice display no ACAT activity (46,47). Therefore, in order to directly determine if ACAT activity was required for the induction of apoptosis by
oxLDL and 7KC we examined the ability of oxLDL and 7KC to induce apoptosis in primary macrophages isolated from ACAT-1 (-/-) mice. **Fig. 3** shows a typical result from TUNEL analysis of MPMs incubated for 24 h with either 50 µg/ml oxLDL or 10 µg/ml 7KC. In comparison to similarly treated wild type MPMs, ACAT-1 (-/-) MPMs display a significantly lower percentage of apoptotic (TUNEL positive) cells (~2.4 fold lower for both oxLDL and 7KC treated cells). Similar results were obtained with 25-OHC treated ACAT-1 (-/-) MPMs (data not shown). Although the absolute value for TUNEL activity varied somewhat among repeat experiments we always found a substantially reduced level of apoptotic cells in ACAT-1 (-/-) macrophages in response to 7KC or oxLDL as compared to control macrophages. These TUNEL assay results were confirmed by performing a caspase-3 activity assay to quantitate the apoptotic induction produced by 7KC or oxLDL treatment of MPM’s isolated and pooled from a large cohort (n>6) of wild type and ACAT-1 (-/-) mice (data not shown).

A cholesterol trafficking inhibitor, U18666A, preferentially inhibits cholesteryl ester accumulation, but does not inhibit 7KC-induced apoptosis.

To address the issue that the apoptotic signal generated by ACAT might result from either the production of cholesteryl esters (CE) or 7KC esters, we took advantage of the observation that oxysterol trafficking occurs at a much faster rate than cholesterol trafficking and may involve a different trafficking mechanism (48). Amphipathic amines, like U18666A, block cholesterol esterification by inhibiting cholesterol trafficking without having a direct effect on ACAT activity (49,50,51,52). Therefore, we determined if U18666A could preferentially inhibit CE formation and, if so, what effect this inhibition would have on the 7KC-induced apoptosis. Using tracer amounts of [14C]cholesterol and [3H]7KC, the accumulation of both
[14C]cholesteryl and [3H]7KC esters in P388D1 cells was observed in response to treatment with unlabeled 7KC (Fig. 4). U18666A limited the accumulation of [14C]cholesteryl esters in 7KC treated cells to levels equivalent to the untreated controls (Fig. 4A). However, U18666A only had a small, but significant ($P = 0.048$), affect on the accumulation of [3H]7KC esters (Fig. 4B). Thus, under these conditions, U18666A differentially inhibited the synthesis of CE over 7KC esters. However, U18666A had no effect on the ability of 7KC to induce caspase-3 activity in P388D1 cells (Fig. 4C).

A phospholipase A$_2$ inhibitor, a global inhibitor of arachidonate metabolism, and an ACAT inhibitor prevent esterification of 7KC and induction of apoptosis by 7KC.

Our previously published results demonstrated that an inhibitor of cPLA$_2$, (AACOCF$_3$) and an inhibitor of AA metabolism (ETYA) prevent the induction of apoptosis by oxysterols (13,34). Therefore, we examined the effect of AACOCF$_3$ and ETYA on the induction of 7KC esters in P388D1 cells. In agreement with the results shown in figure 4, treatment of P388D1 cells with unlabeled 7KC resulted in the formation of [3H]7KC esters from [3H]7KC (Fig. 5). When supplemented with either AACOCF$_3$ or ETYA the stimulation of 7KC ester formation was prevented. In contrast, supplementation with cyclooxygenase inhibitors, FR122047 and NS-398, which do not protect macrophages from 7KC-induced apoptosis (data not shown) were without effect on the induction of [3H]7KC esters (Fig. 5B). Thus, compounds capable of inhibiting 7KC-induced apoptosis in macrophages also prevent the formation of 7KC esters.
Analysis of 7-ketocholesteryl ester content of P388D1 cells by tandem LC/MS.

A range of different oxysterol esters have been shown to accumulate in macrophages upon uptake of modified LDL or 7KC-enriched acetylated LDL (53). To identify the distribution of 7KC esters formed in P388D1 cells, we synthesized authentic standards for the five predominant 7KC esters previously identified in J774.1 macrophages loaded with 7KC-enriched acetylated LDL (53). We then performed a semiquantitative mass spectral analysis of 7-ketocholesteryl esters formed in P388D1 cells after treatment with 7KC for 16 h (Table 1). Macrophage 7KC esters contain primarily saturated and monounsaturated fatty acids, the majority being derived mainly from palmitate and oleate which represented ~21% and ~71% of the total 7KC esters, respectively. Stearate, arachidonate and linoleate made up less than 10% of the total 7KC esters combined. In addition to the five 7KC esters for which we synthesized authentic standards, we also were able to detect 7KC esters of C16:1, C18:2, C20:0 and C20:1 (Fig. 6). Since we did not have standards for these 7KC esters, we were unable to quantitate the relative amounts of each, however, C20:0 and C20:1 were probably also present in significant amounts (~5 – 10% of total).

Arachidonate supplementation can overcome the inhibitory effect of AACOCF₃ on 7KC-induced apoptosis.

The cPLA₂ inhibitor, AACOCF₃, suppresses apoptosis induced by 25-OHC and 7KC (34 and data not shown) as well as the esterification of these oxysterols (Fig. 5 and data not shown). Supplementation of free fatty acids to macrophages in the presence of cPLA₂ inhibition has been shown to restore oxLDL-induced cholesterol esterification (35). Therefore, we determined if supplementation of free fatty acids in the presence of cPLA₂ inhibition could
restore 7KC esterification and if so, what effect this would have on the induction of apoptosis in P388D1 cells. **Fig. 7A** shows that supplementation of P388D1 cells with 50 µM oleic acid or arachidonic acid overcame the inhibitory effect of AACOCF₃ on 7KC-induced formation of [³H]7KC esters. However, only supplementation with arachidonic acid, but not oleic acid, reversed the effect of AACOCF₃ on 7KC-induced caspase-3 activity (**Fig. 7B**). When the cells were further supplemented with the ACAT inhibitor, 58-035, the stimulation of [³H]7KC ester formation produced by arachidonic acid supplementation in the presence of AACOCF₃ was prevented as was the restoration of caspase-3 activity (**Fig. 7A and B**).
DISCUSSION

The current study was initiated to investigate the potential role of ACAT in oxLDL/oxysterol-induced apoptosis in macrophages. The data presented demonstrate that a pharmacological inhibitor of ACAT, 58-035, produces a near complete loss of 7KC-induced apoptosis in a macrophage cell line, P388D1 (Fig. 1), and isolated primary macrophages (Fig. 2A). This suggests that ACAT activity is required for 7KC-induced apoptosis in macrophages. In support of this conclusion, we observed a statistically significant reduction in 7KC-induced apoptosis in ACAT-1 (-/-) MPMs compared to wild type MPMs (Fig. 3). These results are also consistent with our prior observation that oxysterols must be internalized by cells in order to be cytotoxic; even though their initial target is a cell surface calcium channel (11). Based on our previous and currently reported observations, we hypothesize that 7KC not only initiates an apoptotic signaling pathway in macrophages via activation of cPLA₂, but also forms part of a second message in that pathway when esterified by ACAT.

There have been prior indications in the literature that the ACAT may play a role in the cytotoxicity of oxysterols, albeit under somewhat different conditions. Prior reports from two laboratories have described the isolation of ACAT deficient CHO-K1 mutants with selection for resistance to killing by 25-OHC (54,55). One of these studies also described partial resistance to 25-OHC cytotoxicity in wild type CHO-K1 cells treated with 58-035 (54). The selections for 25-OHC resistance were under different conditions than those described for induction of apoptosis in CHO-K1, or other mammalian cells, in that the culture media were cholesterol deficient. These conditions select for regulatory mutations in cholesterol synthesis that act to confer resistance to the down-regulatory effect of 25-OHC on the transcriptional control of cholesterol biosynthetic genes (56). It was suggested, in these prior studies, that the
selection for ACAT deficiency was a cholesterol sparing mechanism since oxysterols are known to stimulate cholesterol esterification in whole cells. However, it is also possible that some of the resistance to killing by 25-OHC, conferred by loss of ACAT or treatment with 58-035, could have been due to inhibition of oxysteryl ester formation.

Oxysterols are important cytotoxic components of oxLDL (17,18) and we present evidence that these oxysterols must be esterified in order to manifest their cytotoxicity. The ability of 58-035 to significantly suppress oxLDL-induced apoptosis in MPMs (Fig. 2B) and the similar observation of attenuated apoptotic induction by oxLDL in ACAT-1 (-/-) MPMs (Fig. 3) are consistent with the hypothesis that the oxysterol components of oxLDL contribute to its cytotoxicity. The residual induction of apoptosis by oxLDL in ACAT-1 (-/-) MPMs and MPMs treated with 58-035 probably reflects the presence of components in oxLDL that are capable of inducing apoptosis by mechanisms independent of ACAT activity (17). In prior studies demonstrating a role for cPLA2 in oxLDL-induced apoptosis we observed a similar attenuation, rather than elimination, of apoptotic induction by oxLDL in MPMs isolated from cPLA2 (-/-) mice (13). In these MPMs the cPLA2-released fatty acid utilized for oxysterol esterification, and according to one report (35), cholesterol esterification, would not be available. Furthermore, it is likely that some of the apoptosis induced by oxLDL, observed in both cPLA2 (-/-) and ACAT-1 (-/-) MPMs, is due to the accumulation of intracellular free cholesterol (57).

We cannot rule out a role for ACAT-2, in addition to ACAT-1, in the oxLDL and 7KC induction of apoptosis in the ACAT-1 (-/-) macrophages (Fig.3). Although Sakashita et al. (58) reported that ACAT-2 is not expressed in resident MPMs; this same report shows that ACAT-2 can be induced during foam cell formation, which does occur when MPMs are incubated with oxLDL. This ambiguity does not exist for our studies with the ACAT inhibitor since it is used
under conditions in which all cholesterol esterification is blocked. In these experiments, we observed a statistical 100% inhibition of 7KC-induced apoptosis in MPMs treated with 58-035 (Fig. 2A) as compared to an approximate 50% inhibition in the ACAT-1 (-/-) macrophages (Fig.3).

It has been suggested that induction of apoptosis by oxysterols and free cholesterol loading proceed through different mechanisms (59,60). The results of the current study agree with this assessment. The in vitro system used to induce apoptosis by free cholesterol loading in macrophages requires the inhibition of cholesterol esterification by treatment with 58-035 (61). In the current study, 58-035 proved to be a potent inhibitor of the induction of apoptosis by 7KC and oxLDL (Fig. 1 and 2). Furthermore, the induction of apoptosis by free cholesterol loading is sensitive to U18666A (60,62) while 7KC-induced apoptosis is insensitive to U18666A (Fig. 4).

Our prior studies have indicated that an arachidonate-derived molecule plays a role in the apoptotic response to oxysterols (13). We now hypothesize that this arachidonate derivative is an arachidonyl oxysterol. Consistent with this hypothesis, the results presented in Fig. 7 show that supplementation of macrophages in the presence of a cPLA2 inhibitor with arachidonic acid, but not oleic acid, restores induction of apoptosis by 7KC. This suggests that it is not the accumulation of 7KC esters in general that produces the apoptotic response, but rather the formation of a specific oxysteryl ester and that 7KC-arachidonate may be a candidate for the apoptosis inducing molecule. Supporting this possibility, mass spectral analysis revealed that P388D1 cells supplemented with 7KC do accumulate 7KC-arachidonate (Fig. 6 and Table 1). Furthermore, Brown et al. (53) detected 7KC-arachidonate among the 7KC esters formed upon loading of MPMs and J774A.1 macrophages with oxLDL.
Our present results also provide an explanation for the observation that oxysterol toxicity can be quenched by cholesterol (63) or a mixture of nontoxic oxysterols (64). Since oxysterols as well as cholesterol are substrates for ACAT (36,37), competition among oxysterols and cholesterol, at the level of ACAT, would be expected to diminish the levels of any specific apoptosis-inducing oxysterol arachidonate generated under these conditions. If, and by what mechanism, such an arachidonyl ester can induce apoptosis in macrophages is not known and is currently under investigation.

The data presented in the current report has potentially important implications in atherogenesis. Our prior results (4) showed that the mitochondrial apoptosis pathway is anti-atherogenic. This suggests that ACAT-1, via its ability to induce apoptosis in macrophages in response to uptake of oxLDL, should be anti-atherogenic. This hypothesis is supported by studies showing that loss of macrophage ACAT, by pharmacological inhibition (65) or by knockout of ACAT-1 (40), results in increased plaque size in animal models of atherosclerosis. In the latter study, an increase in apoptotic cells was observed in the ACAT-1 (-/-) lesions. It was suggested that these apoptotic cells were probably macrophages undergoing cell death in response to free cholesterol toxicity. Therefore, at present it is not possible to make a generic statement concerning the atherogenicity of apoptotic lesional macrophages.

The anti-atherogenicity of macrophage ACAT-1 expression has been attributed to the ability of ACAT to limit the accumulation of free cholesterol. The data presented in this report suggests another function of ACAT-1 is to produce an apoptotic signal in response to oxLDL/oxysterols. Thus, ACAT appears to play a central role in atherogenesis as a regulator of macrophage cell death by reducing accumulation of cytotoxic free cholesterol and also promoting apoptosis in response to oxLDL/oxysterols. Further delineation of the various cell
death pathways followed by in vivo testing of macrophages genetically modified in these pathways will be required to determine the precise roles of macrophage apoptosis in atherosclerosis.

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REFERENCES


**FIGURE LEGENDS**

**Fig. 1.** An ACAT inhibitor prevents 7KC-induced apoptosis in P388D1 cells.

P388D1 cells were preincubated for 1 h with an ACAT inhibitor, 58-035 (10 µg/ml), prior to a 16 h incubation with and without 7KC (10 µg/ml) as indicated in the figure. The cells were then assayed for caspase-3 activity as described in Material and Methods. The data represent the mean ± S.D. of triplicate treatments. *denotes $P < 0.05$ versus 7KC-treated control.

**Fig. 2.** An ACAT inhibitor reduces the induction of apoptosis by 7KC and oxLDL in isolated peritoneal macrophages. The effect of 58-035 on the induction of TUNEL positive cells following a 24 h treatment with 7KC treatment is shown in (A). Resident peritoneal macrophages were isolated, plated on glass coverslips and allowed to attach. The media was changed and supplemented with or without 58-035 (10µg/ml) for 1 h prior to supplementation with and without 7KC (10µg/ml) as indicated in the figure. The incubations were continued for 24 h. Apoptosis was then assayed by in situ TUNEL analysis as described in Materials and Methods. The effect of 58-035 on the induction of TUNEL positive cells following 24 h and 48 h incubations with oxLDL is shown in (B). Isolated resident peritoneal macrophages were supplemented with and without 58-035 (10 µg/ml) for 1 h prior to the addition of oxLDL (50µg/ml). The incubation was continued for 24 h and a 48 h as indicated in the figure. The induction of apoptosis was determined by in situ TUNEL analysis. The results are presented as the mean.
percentage of TUNEL positive cells ± S.D. in ten random fields for each condition. The results shown are representative of three independent experiments. *denotes $P < 0.01$ and **denotes $P < 0.001$ versus 7KC or oxLDL-treated control.

**Fig. 3.** The induction of apoptosis by oxLDL and 7KC is suppressed in ACAT-1 deficient peritoneal macrophages. Resident peritoneal macrophages from wild type and ACAT-1 deficient mice were isolated and treated for 24 h with either oxLDL (50 µg/ml), 7KC (10 µg/ml), or vehicle as indicated in the figure and then subjected to *in situ* TUNEL analysis as described for figure 2. The results shown are representative of three independent experiments. *denotes $P < 0.001$ and **denotes $P < 0.0005$ versus oxLDL and 7KC treated controls, respectively.

**Fig. 4.** Treatment of P388D1 cells with an inhibitor of cholesterol trafficking, U18666A, preferentially inhibits cholesteryl ester accumulation, but does not inhibit 7KC-induced apoptosis. P388D1 cells were supplemented with or without U18666A (5 µM) for 1 h prior to supplementation with and without 7KC (10 µg/ml) as indicated in the figure. At this time, either $[^{14}\text{C}]$cholesterol (0.5 µCi/ml) or $[^{3}\text{H}]$7KC (1.0 µCi/ml) was added to each well. Following a 16 h incubation, total lipids were extracted, and the radioactivity incorporated into cholesteryl esters and 7KC esters was determined (A and B, respectively). The effect of U18666A on the induction of caspase-3 activity in P388D1 cells is shown in (C). P388D1 cells were preincubated with U18666A (5 µM) for 1h prior to a 16 h treatment
with and without 7KC (10 µg/ml) as indicated in the figure. The cells were then harvested and caspase-3 activity was determined as described above. *denotes \( P < 0.05 \) and **denotes \( P = 0.048 \) versus oxysterol-treated control.

**Fig. 5.** 58-035, ETYA, and AACOCF\(_3\) inhibit the formation of 7KC esters in P388D1 cells. The effects of 58-035, ETYA, and AACOCF\(_3\) on the accumulation of \(^{3}\text{H}\)7KC esters in P388D1 cells (A). P338D1 cells were cultured in the presence of \(^{3}\text{H}\)7KC (1.0 µCi/ml) in media supplemented with either 58-035 (10 µg/ml), ETYA (20 µM), AACOCF\(_3\) (20 µM) as indicated in the figure. Unlabeled 7KC (10 µg/ml) was added and the cells were incubated for 16 h. Total lipids were extracted and the radioactivity incorporated into 7KC esters was determined. The effects of a COX-1 inhibitor, FR122047 (10 µM) and a COX-2 inhibitor, NS-398 (5 µM) on the accumulation of \(^{3}\text{H}\)7KC esters in P388D1 cells as compared to treatment with 58-035 and ETYA (B). P388D1 cells were cultured in the presence or absence of the indicated inhibitors for 1 h prior to supplementation with \(^{3}\text{H}\)7KC and unlabeled 7KC for 16h as describe for panel A. *denotes \( P < 0.05 \), **denotes \( P = 0.12 \), and ***denotes \( P =0.33 \) versus oxysterol-treated control.

**Fig. 6.** SRM chromatographic traces of a standard mixture (upper figure) containing 16:0, 18:0, 18:1, 18:2 and 20:4 7KC esters, as described in the Methods and Materials. SRM traces of a macrophage total lipid extract (bottom figure) represents analysis of about 1.4% of the total extract. The top panel in
each figure contains overlaid SRM traces for 16:0 and 16:1, the middle panel contains traces for 18:0, 18:1, 18:2 and 18:3, and the bottom panel contains traces for 20:0, 20:1 and 20:4. The amplification factors for each trace relative to 7KC-18:0 are shown in parentheses; the same factor was applied to each SRM trace in a panel.

**Fig. 7.** The effect of supplementing arachidonic acid and oleic acid in the presence of AACOCF₃ on the formation of [³H]7KC esters and the induction of caspase-3 activity by 7KC. The effect of arachidonic acid or oleic acid supplementation on AACOCF₃ inhibition of [³H]7KC ester formation is shown in (A). P388D1 cells were cultured in the presence or absence of AACOCF₃ (20 μM) or 58-035 (10 μg/ml) for 1 h and then supplemented with and without 7KC (10 μg/ml) and either oleic acid or arachidonic acid (50 μM) complexed to fatty acid free BSA as indicated in the figure. [³H]7KC (1.0 μCi/ml) was added to the cells and following a 6h incubation, the lipids were extracted and the radioactivity incorporated into [³H]7KC esters was determined as described in figure 5. The effect of arachidonic acid or oleic acid supplementation on AACOCF₃ inhibition of 7KC-induced apoptosis is shown in (B). P388D1 cells were supplemented with and without oleic acid or arachidonic acid (50 μM) in the presence of either AACOCF₃ (20 μM) or 58-035 (10 μg/ml) for 1 h prior to the addition of unlabeled 7KC (10 μg/ml) as indicated in the figure. Following a 16 h incubation, the cells were harvested and caspase-3 activity was determined as described above.
Table 1. 7-ketocholesteryl ester content of P388D1 cells.

<table>
<thead>
<tr>
<th>7-Ketocholesteryl Ester¹</th>
<th>Normalized Amount (%)</th>
<th>pmol/mg cell protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>7-Ketocholesteryl palmitate (16:0)</td>
<td>20.98 ± 0.36</td>
<td>66.42 ± 7.15</td>
</tr>
<tr>
<td>7-Ketocholesteryl stearate (18:0)</td>
<td>7.26 ± 0.88</td>
<td>22.81 ± 1.18</td>
</tr>
<tr>
<td>7-Ketocholesteryl oleate (18:1)</td>
<td>71.12 ± 1.16</td>
<td>225.26 ± 25.57</td>
</tr>
<tr>
<td>7-Ketocholesteryl linoleate (18:2)</td>
<td>0.22 ± 0.13</td>
<td>0.71 ± 0.51</td>
</tr>
<tr>
<td>7-Ketocholesteryl arachidonate (20:4)</td>
<td>0.43 ± 0.13</td>
<td>1.35 ± 0.45</td>
</tr>
</tbody>
</table>

¹The numbers in parentheses refer to the length of the carbon chain and number of double bonds of the fatty acyl chain.

The MS response factors were calculated as described in the Materials and Methods.

P388D1 cells were incubated for 16 h in media supplemented with 20 µg/ml 7KC. The cells were rinsed twice with PBS and total lipids were extracted for mass spectral analysis as described in Materials and Methods. Data provided are of individual 7KC esters expressed as a percentage of the total measured oxysteryl ester pool in the cell and as the absolute masses of the detected oxysteryl ester expressed as pmol/mg cell protein. All data are means ± S.D. of triplicate samples.
Fig. 1

Caspase-3 Activity (FLU/mg x106)

Control 58-035

Vehicle + 7KC

*
Fig. 2

A.  % TUNEL Positive Cells

- Control
- 58-035

B.  % TUNEL Positive Cells

- 24 hour
- 48 hour

Vehicle  + oxLDL  + oxLDL + 58-035

- Vehicle
- + 7KC
Fig. 3
Fig. 4

A. [\textsuperscript{14}C]Cholesterol esters (dpm/mg $\times 10^5$)

- Control
- U18666A
- Vehicle
- + 7KC

B. \[^3\text{H}]\text{7KC esters} (dpm/mg $\times 10^6$)

- Control
- U18666A
- Vehicle
- + 7KC

C. Caspase-3 Activity (FLU/mg $\times 10^6$)

- Control
- U18666A
- Vehicle
- + 7KC
Fig. 5

A. FR-122047

B. Vehicle

$[^3]H$7KC esters
(dpm/mg $\times 10^6$)

Control

58-035

ETYA

AACOCF

NS-398

FR-122047

Vehicle + 7KC

$^*$

$^{**}$

$^{***}$

$^*$

$^{**}$

$^{***}$
Fig. 6

![Graph showing the relative abundance of different lipid classes over time. The graphs display peaks at specific times and relative abundances for 7KC-16:x, 7KC-18:x, and 7KC-20:x.](image-url)