Antipsychotic-induced increase in lipid biosynthesis – activation through inhibition?

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Antipsychotic drugs have become indispensable in the treatment of schizophrenia since the introduction of first-generation antipsychotics (FGAs; e.g., chlorpromazine and haloperidol) in the 1950s. Unfortunately, several of the commonly used drugs, particularly the second-generation antipsychotics (SGAs) clozapine and olanzapine, induce metabolic disturbances such as obesity, hypertriglyceridemia, glucose dysregulation, and, in some studies, elevated serum cholesterol levels (1, 2). These adverse effects represent major challenges in the treatment of psychotic disorders, as they reduce compliance and contribute to increased cardiovascular mortality among patients (3). Despite these problems, the use of metabolically potent antipsychotic drugs is widespread, arguably due to superior therapeutic efficacy compared to antipsychotics with more favourable metabolic profiles (4).

Antipsychotic effect is related to blockage of dopamine D2 receptors in the brain. The molecular mechanisms mediating metabolic disturbances are incompletely understood. While antagonistic effects on serotonin 5HT2C and histamine H1 receptors in the hypothalamus seem to be relevant for antipsychotic-induced weight gain through stimulation of appetite (5), other mechanisms are also likely to contribute. In 1965, haloperidol was demonstrated to inhibit cholesterol biosynthesis (6). Later on, it has been shown that other cationic amphiphilic drugs, like the FGA chlorpromazine and the antidepressant imipramine, can increase lysosomal content of cholesterol and interfere with the sterol regulatory element binding protein (SREBP)-mediated cholesterol-sensing system in the endoplasmic reticulum (ER) (7-10). We and others, however, have demonstrated that antipsychotic drugs induce transcriptional activation of cholesterol and fatty acid biosynthesis genes controlled by the SREBP1 and SREBP2 transcription factors (11, 12). How these apparently divergent in vitro findings may be unified to form a common theory has not yet been sorted out, particularly with respect to their possible role in explaining antipsychotic-induced metabolic adverse effects in the clinical setting.

In this issue of *Journal of Lipid Research*, Canfrán-Duque et al. (13) contribute new, interesting data on the effect of antipsychotic drugs on cellular cholesterol biosynthesis and transport, elaborating previous work by others and themselves (6, 14-16). They demonstrate that the FGA haloperidol and the SGAs clozapine, risperidone, and ziprasidone reduce de novo cholesterol biosynthesis in three different types of cultured human cells (HepG2, SH-SY5Y and HL-60). This occurs through inhibition of several enzymatic steps in the later part of the cholesterol biosynthesis pathway, such as Δ^7^-reductase, Δ^5,7^-isomerase and Δ^14^- reductase, leading to accumulation of various cholesterol precursors. The resultant reduction in cholesterol biosynthesis may seem paradoxical in light of the well-established antipsychotic-related SREBP-controlled activation of lipid biosynthesis genes (11, 17). However, the authors suggest that transcriptional activation of cellular lipogenesis is, in fact, a homeostatic feedback mechanism triggered by reduced cholesterol biosynthesis, which is reinforced by antipsychotic-induced trapping of LDL-derived cholesterol within the endosomes/lysosomes, confirming formerly published results (16). In contrast to the reduced cholesterol production, biosynthesis of complex lipids (triglycerides and phospholipids) was increased in the cell cultures during exposure to antipsychotic drugs. The authors suggest that these opposite effects are indeed linked, since enzymes involved in triglyceride biosynthesis are not inhibited by the antipsychotics. The compensatory SREBP activation thus leads to elevated biosynthesis of free fatty acids, triglycerides, and phospholipids, as outlined in Figure 1.
Of note, the propensity of the individual antipsychotics to stimulate lipid biosynthesis changed under washout conditions, where growth medium containing antipsychotic drugs was replaced with drug-free medium for 4 hours before cellular lipid concentrations were measured. In these experiments, clozapine came out as the most potent stimulator of fatty acid, triglyceride, and phospholipid biosynthesis. Removal of antipsychotics from the culturing medium also led to induction of cholesterol production, in agreement with previous findings (18). These data demonstrate how modest changes in experimental setup may lead to different conclusions, emphasizing the importance of careful planning of experimental design in future studies.

The in vivo significance of inhibited cholesterol biosynthesis as a causative mechanism for antipsychotic-induced triglyceride biosynthesis remains to be determined. Studies in rats have demonstrated that antipsychotic drugs indeed stimulate SREBP1-controlled gene expression and elevate lipid levels, both in acute (19, 20) and in subchronic experiments (21, 22). Although the washout conditions applied in the cell culture experiments may seem somewhat artificial, it should be noted that the half-life of antipsychotic drugs in rats is very short, resulting in oscillation of serum drug concentrations when the standard procedure of administering antipsychotics twice daily is followed. In female rats, increased lipogenic gene expression and elevated serum triglycerides were found 20 hours after the last drug dose, when serum concentrations were negligible, implying that a washout-like effects may have contributed to lipogenic activation in these conditions (22). In patients, however, serum levels of antipsychotics reach steady-state conditions after approximately 1 week at stable doses (23) and a direct parallel to the washout scenario observed in cell cultures seems less clear.

Among the antipsychotics examined, Canfrán-Duque et al. found that the FGA haloperidol and the SGAs risperidone and ziprasidone targeted the same enzymes in their inhibition of cholesterol biosynthesis, although with different relative activities: ziprasidone > haloperidol > risperidone. In contrast, clozapine mainly affected Δ24- reductase and Δ8,7-isomerase activity, although with weaker effects than the other antipsychotics examined. Clozapine is clearly the most metabolically unfavourable drug in patients, while ziprasidone and haloperidol have minor effects on serum lipids and other metabolic parameters (1, 24, 25). It seems puzzling, therefore, that depletion of cholesterol (with subsequent increase in triglyceride biosynthesis) in the cells was most pronounced during exposure to ziprasidone, with limited effects of clozapine. Whether the difference in enzymatic targets and degree of inhibition may be relevant with regard to this paradox remains to be elucidated. In this regard, it would be very interesting to include olanzapine in the group of drugs examined, since this antipsychotic is both highly metabolically potent and frequently used.

In summary, the findings by Canfrán-Duque and colleagues provide interesting new insight into the effects of antipsychotic drugs on lipid biosynthesis in vitro. The authors launch a new, integrative theory on how inhibition of cholesterol biosynthesis may be relevant for the antipsychotic-induced increase in serum triglyceride levels observed both in rodent models and in patients. Once the basic mechanisms have been firmly established, it is time to investigate the role of antipsychotic-induced inhibition of cellular cholesterol production and homeostatic lipogenic activation in dyslipidemia and obesity.
References:


Figure 1: Cellular lipid homeostasis and regulation of SREBP-controlled cholesterol and fatty acid biosynthetic gene expression, in the absence or presence of antipsychotic drugs.
