Atherosclerosis (AS), a major etiology of cardiovascular disease, is considered to be a chronic inflammatory disease characterized by excessive inflammatory cells, such as macrophages, accumulated in the arterial wall (1). As the main effector cells of the immune/inflammatory system, macrophages engulf lipids and produce various inflammatory factors, thus participating in the progress of AS (1–3). Therefore, it is very important to clarify the mechanisms that regulate macrophage-related inflammatory response for the prevention of AS.

The report in the Journal of Lipid Research by Schneider et al. (4) shows that apolipoprotein A-I binding protein (AIBP), a secreted protein that avidly binds to apoA-I, the major component of HDL, plays a key role in regulating macrophage cholesterol efflux and inflammation in AS (5). ApoA1bp−/−/Ldlr−/− mice fed a high-fat diet have shown exacerbated hypercholesterolemia, hypertriglyceridemia, and larger atherosclerotic lesions compared with Ldlr−/− mice. Conversely, overexpression or injection of AIBP reduced aortic inflammation and atherosclerotic plaques. In vitro, AIBP facilitated cholesterol efflux, the first step of reverse cholesterol transport, from cultured macrophages to HDL, reducing the cholesterol content in lipid rafts that inhibited the inflammatory responses to lipopolysaccharide (LPS), suggesting the crucial role of cholesterol efflux on AIBP-mediated anti-inflammatory and immunosuppressive functions.

Previous research demonstrated that AIBP promotes ABCA1-mediated cholesterol efflux from endothelial cells and macrophages via interaction with apoA-I (6–8). In zebrafish and mice, AIBP regulated cholesterol levels in endothelial cells that control angiogenesis via depleting lipid raft content in the cell membrane, therefore inhibiting vascular endothelial growth factor receptor 2 and upregulating Notch signaling (6, 8). Meanwhile, AIBP-mediated cholesterol efflux can also impair the lipid raft-containing Toll-like receptor 4 (TLR4), which has been shown to upregulate the MYD88-mediated activation of MAPK and NF-κB signaling as well as downstream inflammatory cytokines (9, 10). In addition, AIBP has been found to promote the binding of apoA-I to ABCA1 in macrophages and prevent ABCA1 protein from COP9 signalosome subunit 2-mediated degradation so as to prevent foam cell formation (7).

Several studies have revealed that ABCA1 can directly function as an anti-inflammatory receptor for apoA-I to suppress inflammation independently of its cholesterol efflux activity (11,12). The interaction of apoA-I and ABCA1 stimulated the JAK2/STAT3 signal pathway and tristetraprolin-dependent posttranscriptional regulation of pro-inflammatory cytokines mRNA decay (11). Knockout of ABCA1 in mice increases inflammatory cell infiltration in a number of tissues, including the vessel wall and peritoneal cavity, and blood circulation (13). These results suggest that secretory AIBP modulated the immune/inflammatory response through regulating lipid transport and lipid raft-related receptor activity.

Interestingly, there may be more mechanisms to explain the action of AIBP on inflammation and AS. The Apoa1bp and the NAXE gene that codes AIBP was renamed as the NADHX epimerase (NAXE) by the Human Gene Nomenclature Committee. As an epimerase in mitochondria, AIBP converts R-NADHX, R-epimers of nicotinamide adenine dinucleotide hydration (NADHX), to biologically useful S-NADHX that rapidly reconverted to nicotinamide adenine dinucleotide (NADH) (14). NADHX has been shown to inhibit several dehydrogenases (15), which is necessary for mitochondria oxidative phosphorylation (OXPHOS) (16). OXPHOS, the major function of mitochondria, has recently emerged as a central organelle that integrates cellular metabolism and inflammatory responses (17). Yu et al. (18) have recently found that OXPHOS is reduced in AS, promoting mitochondrial

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dysfunction and necrotic core formation. The damaged mitochondria accumulated in macrophages results in the activation of the NLRP3 inflammasome and production of IL-1β (19). In addition, impaired mitochondrial OXPHOS has been found to prevent the repolarization of pro-inflammatory macrophages to anti-inflammatory macrophages (20). Recently, mutations of NAXE in children have been reported to result in acute-onset ataxia, cerebellar edema, spinal myelopathy, and skin lesions. Increased lactate in cerebrospinal fluid and R-NADHX in fibroblasts have been shown in these diseases, indicating that NAXE is an unheeded target for controlling metabolism and the immune/inflammation system (21).

These studies improve our understanding of the potent anti-inflammatory properties of extracellular and intracellular AIBP (Fig. 1). Serum AIBP, mainly secreted from liver and kidney, inhibits inflammatory cytokine expression via TLR-4/MyD88-mediated MAPK and NF-κB signaling as well as the ABCA1/JAK2/STAT3 pathway after binding with apoA-I. On the other hand, mitochondrial AIBP may function as an NADHX epimerase to prevent OXPHOS damage and mitochondrial dysfunction (Fig. 1). Moreover, lipid and energy metabolism may also be involved in the process of AIBP-mediated anti-inflammatory response, and this suggests that AIBP may be a novel therapeutic target for chronic metabolic inflammatory disease, such as AS.

**REFERENCES**


